

AreTomo2 1.1 User Manual

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AreTomo2, an open-source version of **AreTomo 1.4.3**, is a multi-GPU accelerated software package that enables fully-automated marker-free tomographic alignment and reconstruction (Zheng et al., 2022). The alignment scheme corrects various defects arising from the data collection including translational misalignment, in-plane rotation, tilt-angle offset, and anisotropic 3D local motion due to beam induced motion. Both weighted back projection (WBP) and simultaneous algebraic reconstruction (SART) are implemented and integrated with tomographic alignment. Together with the recent addition of CTF estimation, AreTomo2 is capable of providing, in a single application, not only the tomograms but also the crucial alignment parameters needed to bootstrap subtomogram averaging workflow. AreTomo2 can be easily configured via command line parameters to suit various practical needs. It is our best wish that this new release can expedite your quests for uncovering structures at high resolutions and promote the advancement and the interest in cryoET.

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1. System Requirement and Installation

AreTomo2 is a multi-GPU accelerated Linux application written in C++/CUDA. A Linux system equipped with one or more advanced NVidia GPU cards is needed. The CPU memory allocated in AreTomo2 is roughly twice the size of the input tilt series calculated based upon 4 bytes per pixel. AreTomo2 is a single-program application. Once unzipped, no further installation is needed if Cuda driver and toolkit have been installed already.

1.1 Install from source

If the source code is downloaded, the binary can be generated using two included **makefile** and **makefile11**. Open a Linux terminal and execute at the command prompt **make exe** if CUDA 10.x is installed or **make exe -f makefile 11** when CUDA 11.x or 12.x is installed. If you do not know what CUDA version is installed, it can be found by running **nvcc -- version** at the command prompt.

Both **makefile** and **makefile11** define CUDAHOME, a variable that holds the full path to CUDA toolkit installation point. Please use CUDAHOME in the provided makefiles as an example only to help you locate where CUDA toolkit is installed on your system and revise the assignment to CUDAHOME accordingly.

1.2 Compile the library files

There are two precompiled library files, **libutil.a** and **libmrcfile.a** in LibSrc/Lib. If you run into any problems in linking these two files, try recompiling them using the contained makefiles. Compile **libutil.a** first followed by **libmrcfile.a**. Run **make clean** and then **make all** in LibSrc/Util directory and this will generate **libutil.a** that is automatically moved into /LibSrc/Lib. Repeat the same steps in LibSrc/Mrcfile.

2. Quick Start

AreTomo2 can be invoked from Linux command line. The version number and built date can be found by running the following command.

AreTomo2 --version

Users can get a quick help information about the command line parameters by running the following command.

AreTomo2 --help

When the above two commands can be executed, it is a good sign that the underlying libraries required by AreTomo2 are properly installed. The following is an example that takes an input tilt series and generates a 3D volume.

AreTomo2 -InMrc tiltSeries.mrc -OutMrc volume.mrc -VolZ 1200 -OutBin 4

As can be seen, AreTomo2 requires all the projection images be stored in a single MRC file. In this example the input MRC file is **tiltSeries.mrc**. The output file, **volume.mrc**, stores the generated 3D volume. **-VolZ 1200** sets the volume height or the z dimension to 1200 unbinned voxels. **-OutBin 4** causes the generated volume to be 4x binned before it is saved to disk. Assuming 3838x3710 pixels per

tilt image, the unbinned volume has 3838x3710x1200 voxels. **-OutBin 4** reduces the volume to 958x926x300 voxels.

It is important to note that in this example tilt angles are expected to be stored as the first float in the extended headers of **tiltSeries.mrc**. If not, users must use other means described in the next section to pass tilt angles into AreTomo2. Otherwise, the program will exit immediately.

3. Input of Tilt Angles

Tilt angles are required input parameters by AreTomo2. When tilt angles are NOT stored in the extended headers, there are two ways for users to pass them into AreTomo2.

3.1. -TiltRange

This option is best suited for a tilt series collected with a fixed angular step and tilt images in the MRC file are sorted according to their tilt angles. **-TiltRange** should be followed by two end angles as shown in the following example.

AreTomo2 -InMrc tiltSeries.mrc -OutMrc volume.mrc -VolZ 1200 -OutBin 4 -TiltRange -60 60

-TiltRange -60 60 informs AreTomo2 that **tiltSeries.mrc** is collected in the range of $[-60^\circ, 60^\circ]$ and the tilt images are arranged in ascending order of their tilt angles.

3.2. -AngFile

Tilt angles can be passed into AreTomo2 using a white-space separated two-column text file using **-AngFile**. Here is an example.

AreTomo2 -InMrc tiltSeries.mrc -OutMrc volume.mrc -VolZ 1200 -OutBin 4 -AngFile tiltAngles.txt

The first column lists the tilt angles in the same order as that of tilt images in the input MRC file. The second column lists either 0 or 1-based acquisition indices sorted in chronological order of image acquisition. Although the second column is optional, its absence will disable dose weighting and generation of ordered list files (csv files) needed by Relion subtomo averaging.

Note that in the older versions of AreTomo and AreTomo2, the second column lists the accumulated dose rather than the acquisition index.

4. -VolZ

4.1. Control volume thickness

The z axis refers to the one perpendicular to the projection images. As mentioned earlier, **-VolZ** provides a means for users to specify z height of the reconstructed volume. Ideally, we would like to choose a value such that the reconstructed volume just contains the entire structure with neither clipping nor too many empty voxels. In practice, users can estimate the z dimension based upon sample thickness and the pixel size of projection images and then add 200 voxels for precaution. Another option is to run AreTomo2 with an educational guess combined with a large binning for **-OutBin**, say 4 or even 8. Upon completion, review the x-z slice of the reconstruction and make corresponding adjustment based upon what you see. Note again the input unit for volume z height is unbinned voxel.

4.2. Generate aligned tilt series

Aligned tilt series will be saved in the output MRC file when **-VolZ 0** is included in the command. The aligned tilt images are sorted in ascending order of tilt angles in the output MRC file with tilt angles saved in MRC extended headers.

When dose weighting is enabled at the command line, the output is the dose-weighted aligned tilt series. If such a tilt series is intended to be reconstructed in the future, make sure to disable dose weighting at the command line. See section 12 for details.

An angle file is also created in the same directory along with the output MRC file. This file has the same name as the output MRC file but ended with ".tilt". This file can be edited to have a second column for accumulated dose should the aligned tilt series be reconstructed with dose weighting enabled. See section 12 for details.

4.3. Reconstruct aligned tilt series

If users want to reconstruct the aligned tilt series, it can be done by adding **-Align 0** and **-VolZ** followed by a positive integer in the command line. **-Align 0** prevents the alignment process from running on the already aligned tilt series.

5. Input tilt axis

In general, AreTomo2 does not need to know the orientation of tilt axis. However, if there are several projection images bearing large translational shifts, the tilt axis may not be determined accurately. If you are not satisfied with the quality of the reconstructed volume, you may want to try to bootstrap AreTomo2 with **-TiltAxis** followed by the user provided value.

```
AreTomo2 -InMrc tiltSeries.mrc -OutMrc volume.mrc -VolZ 1200 -OutBin 4 -TiltRange -60 60 \
-TiltAxis 10.5
```

Note that the orientation of tilt axis is relative to the y-axis (vertical axis of tilt image) and rotates counter-clockwise.

6. Measure and correct tilt angle offset

Tilt offset refers to the difference between the actual tilt angles and those read from microscope during data collection. This single-value offset arises due to either sample mounting or milling angle. For a slanted sample, a larger z height than the sample thickness is needed for reconstruction without clipping the reconstructed structure, resulting a larger MRC file to store the bigger volume. See Fig. 1 for the effect of the correction of tilt offset.

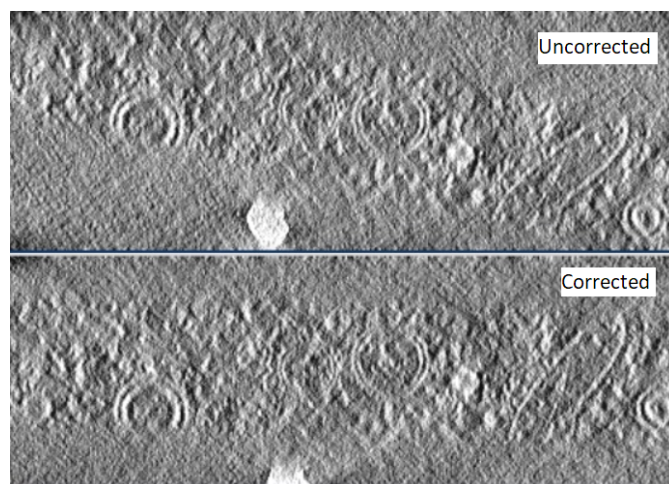


Fig. 1 (Top) Slanted sample reconstructed without the correction of tilt offset. (Bottom) Reconstruction with the correction of tilt offset.

Please note that the orientation of the missing wedge will be changed as a result of the correction of tilt offset. For subtomogram averaging, tomograms reconstructed from tilt series collected within the same tilt range may have different orientations of missing wedges.

-TiltCor can be followed by three values. **-TiltCor 1** results both the measurement and correction of tilt offset. **-TiltCor 0** only measures the tilt offset and is the default setting. The measured offset is corrected only for alignment but not for final reconstruction. **-TiltCor -1** disables the measurement and the correction. Occasionally, the measurement is erroneous and can impair the alignment accuracy.

In the case the tilt offset is not measured properly, an estimated tilt offset can be entered as the second parameter to **-TiltCor**. For example, **-TiltCor 0 10** adds 10° tilt offset to each tilt angle only for the alignment. **-TiltCor 1 10** adds 10° tilt offset to each tilt angle for both alignment and reconstruction. The tilt offset can be estimated by performing a global only alignment and a quick reconstruction using 4x or even higher binning (**-OutBin 4**) followed by reviewing the xz slice.

7. Flip intensity: **-FlipInt**

By default, the reconstructed volume and the input tilt series use the same grayscale that makes dense structures dark. **-FlipInt 1** makes the dense structures white instead.

8. Flip volume: **-FlipVol**

By default, the x-z slices of the reconstructed volume are saved according to their y coordinates in the output MRC file. **-FlipVol 1** saves x-y slices instead according to their z coordinates.

9. Reconstruction schemes

By default, 3D volumes are computed by SART reconstruction. However, weighted back projection can be enabled by setting **-Wbp 1**.

SART is the default reconstruction with the number of iterations and the number of projections per subset set to 20 and 5, respectively. If users want to change these two settings, -Sart can be used for this purpose followed by the number of iterations and number of projections per subset.

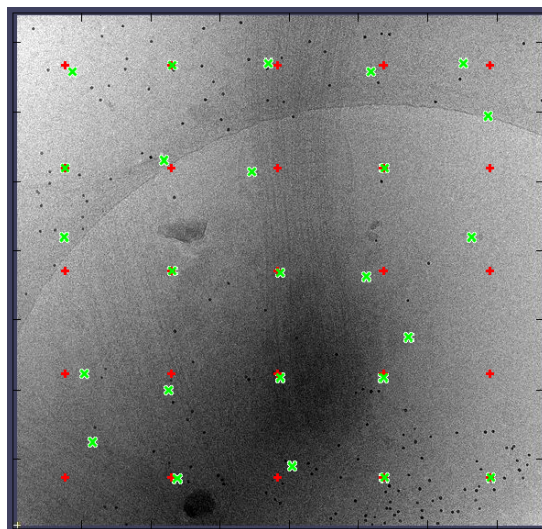


Fig. 2 Uniform and smart grids for local motion measurement and correction. In uniform grid the red-labeled patch centers are evenly distributed as opposed to the green-labeled patch centers in smart grid where patch centers are kept away from empty areas.

10. Local motion correction

Beam induced motion significantly complicates the situation in cryoEM tomography. Specimen movement results from not only stage tilting but also beam induced motion. Tomographic alignment premised only on single-axis rotation, regardless fiducial based or not, are not accurate enough for high-resolution tomography if beam induced local motion is left uncorrected. AreTomo2 provides two means to correct the local motion.

10.1. -Patch

-Patch is followed by two positive integers, e.g. **-Patch 5 4**, that specify number of patches in x and y directions, respectively, where local motions are measured and then corrected. Prior to version 1.2.0, the grid of patches is uniform where the patch centers are evenly distributed. AreTomo2 1.2.0 implements the smart grid that dodges the empty areas for local motion measurement. The uniform and

10.2. Isolated or Sparse Sample: -RoiFile

-RoiFile requires users to provide a list of x and y coordinates and put them in a two-column text file, one column for x and the other for y. Each pair defines a region of interest (ROI). AreTomo2 then performs focused alignment as described in section 11 on the structure in each of these ROIs. Local motion is then interpolated based upon a distance-based scheme and corrected at each pixel of each projection image before 3D reconstruction is performed. **Fig. 2** shows an example where nine targets

were selected along the elongated structure. As mentioned before, the origin of the coordinate system is at the image's lower left corner.

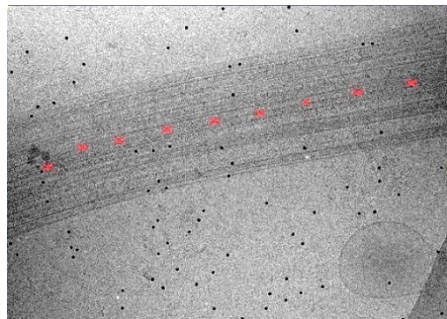


Fig. 3 Illustration of target selection along an elongated structure required to activate local motion correction.

11. Dose Weighting

Once the input tilt series is aligned, dose weighting can be applied to the aligned tilt series before it gets reconstructed. To activate dose weighting, high tension (**-Kv**), pixel size in angstrom (**-PixSize**), dose per tilt in e/A^2 (**-ImgDose**), and the second column of angle file (**-AngFile**) are needed. Here is an example of angle file with the second column with tilt images collected using dose symmetric scheme:

```
-45.01 31
-42.02 28
.....
-0.02 1
.....
41.98 29
44.98 30
```

-Kv 300 -PixSize 3.5 -ImgDose 2.0 -AngFile PathToAngFile/myAngFile.txt

12. Alignment file

AreTomo2 automatically generates a text file containing both global and local alignment parameters. Named after the input MRC file, this file uses **aln** as file extension and is saved in the output directory containing the reconstructed volume. Should users decide to reconstruct the volume with a different setting, the corresponding aln file can be loaded by means of **-AlnFile** to bypass the lengthy alignment process, as is illustrated in the following example.

AreTomo2 -InMrc tiltSeries.mrc -OutMrc ~/Temp/volume.mrc -AlnFile ~/Temp/tiltSeries.aln \

-VolZ 1200 -OutBin 1

13. Volume z height for alignment: -AlignZ

This function specifies the z height of the temporary volume reconstructed for projection matching as part of the alignment process. This value plays an important role in alignment accuracy. The default value is 600. This z height should be always smaller than that specified by **-VolZ**, as illustrated in Fig. 4. As can be seen, the z height for alignment is preferred to be close to the sample thickness whereas the z height for final reconstruction should be larger.

In practice we can perform global only alignment and 3D reconstruction with a large binning. The sample thickness can then be measured in the xz slice of the volume.

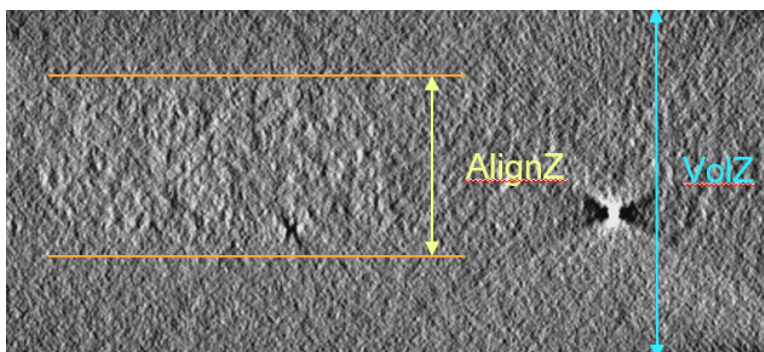


Fig. 4 Illustration of **-AlignZ** and **-VolZ** in a xz slice of a reconstructed volume.

14. Reconstruction without alignment: -Align 0

This option lets users reconstruct tomogram without aligning the input tilt series, which must be, therefore, pre-aligned either by AreTomo2 or some other software. One application of this option is that a raw tilt series is first aligned and then CTF-corrected. The aligned and CTF-corrected tilt series is finally input into AreTomo2 to reconstruct CTF-corrected tomogram.

15. Tilt axis determination: -TiltAxis initVal refineFlag

Tilt axis determination is a two-step processing in AreTomo2. A single tilt axis is first calculated followed by the determination of how tilt axis varies over the entire tilt range. The “initVal” lets users enter their estimate and AreTomo2 refines the estimate in $[-3^\circ, 3^\circ]$. If users do not know, they can enter 0 instead and AreTomo2 will search an optimal value in full range. The “refineFlag” can be one of -1, 0, and 1. The -1 instructs AreTomo2 to the user input value without any refinement. The default setting is 0 and 1.

refineFlag	Meaning
-1	Use the input value without any refinement

0	Refine the input value and use the refined value for the entire tilt series
1	Refine the input value and calculate tilt axis at each tilt angle.

16. Output Imod files for subtomogram averaging

Since version 1.1.0 AreTomo2 can output the files needed for Relion4 and Warp to start subtomogram averaging. The setting -OutImod is used to generate the needed files for this purpose. A subfolder named after the output MRC file is created, if not present, under the output directory. For example, “-OutMrc [MyDir/MyTomo.mrc](#) -OutImod 1” creates the subfolder [MyTomo_Imod](#) in [MyDir](#). The generated files listed in the following table are placed in the subfolder.

xxx.st	MRC file of either aligned or raw tilt series without dark images
xxx.tlt	Single-column text file containing the tilt angles
xxx.xf	Imod transformation file
newst.com	Imod file
tilt.com	Imod file
xxx.csv	ordered list containing acquisition indices and tilt angles.

Note that -OutImod 0 disables this function and does not create the subfolder and the corresponding files and this is the default setting.

16.1 -OutImod 1 (Relion4)

This setting is for Relion4 subtomogram averaging and does not generate [xxx.st](#) file since the original tilt series is used as input. The path of the original tilt series is given in the [newst.com](#).

AreTomo2 may reject the dark images in its alignment process when they are detected, see Section 17 for details. In this case, [tilt.com](#) contains a line (EXCLUDELIST) that lists all the rejected images. To maintain the data consistency across different files, [xxx.xf](#), while containing the transformation matrices for the included images, also have the unit matrices, for the rejected image. [xxx.tlt](#) lists the tilt angles of all images including the rejected ones. The tilt angles are ordered according to the images in the original tilt series.

It is not recommended to use -TiltCor 1 when subtomogram averaging will be performed afterwards. This is because -TiltCore 1 causes tomograms to tilt and changes particle z-coordinates in the tomograms.

The generated csv file has two comma-separated columns. The first column lists 1-based acquisition indices in ascending order and the second is the corresponding tilt angles.

16.2 -OutImod 2 (Warp)

This setting is intended for Warp based subtomogram averaging. It generates a [xxx.st](#) file that stores all the original images except the rejected ones. Correspondingly, [xxx.tlt](#) and [xxx.xf](#) contain only the entries for the included images. [newst.com](#) has a line pointing to the [xxx.st](#) file. [tilt.com](#) does NOT have a line for EXCLUDELIST.

Note that, since the subfolder contains all the files needed for Relion4, this setting is expected to meet the Relion4 requirement, although we have not tested the compatibility.

16.3 -Outlmod 3 (Global- and local-aligned tilt series)

An accurate tomographic alignment is expected to be beneficial for subtomogram averaging, allowing it to start at the vicinity of ground truth. Unfortunately, `xxx.xf` file can only have the transformation matrices of the global alignment, leaving no room for the local alignment parameters determined by AreTomo2. This setting provides a path for subtomogram averaging to take the advantage of AreTomo2's local alignment. To this end, the aligned and dark image-rejected tilt series is generated and saved in the `xxx.st` file. As a result, the `xxx.xf` file is filled with unit transformation matrices to prevent `xxx.st` from being aligned again. Since the interpolation in the generation of the aligned tilt series attenuates the high-frequency information, version 1.3.0 intentionally enhances the high-frequency components of the raw images beforehand to alleviate the attenuation due to interpolation.

16.4 Suggestions for subtomogram averaging

Since AreTomo2 may exclude dark images and adjust tilt angles, it is recommended to use AreTomo2 generated tomograms for particle picking and then feed the needed file in the created subfolder to the subsequent processing pipeline. The mismatch between tilt angles, tilt series, and tomograms, potentially severe for subtomogram average, can be avoided.

It is NOT recommended to use `-TiltCor 1` when subtomogram averaging will be performed afterwards. This is because correction of tilt angle offset rotates the tomogram. As a result, the z coordinates of picked particles deviate from in-sample z depth.

17. Remove dark images

Since version 1.1.0, AreTomo2 allows users to decide how stringent the dark images are detected and removed using `"-DarkTol threshold"`. The default threshold is 0.7. Lowering the threshold means less images are removed. The removed dark images are listed by their indices in a newly created text file named after the input MRC file name and appended with `"_DarkImgs.txt"`.

18. CTF estimation

Since version 1.4.0 AreTomo2 has added a function that estimates the CTF of each tilt images. This function will be enabled when users provide pixel size, high tension, and spherical aberration Cs of the tilt series. The following is an example of the corresponding command line parameters.

AreTomo2 -PixSize 2.4 -Kv 300 -Cs 2.7

If the data is collected with a phase plate, the resulting phase shift can be estimated using the following command line.

AreTomo2 -PixSize 2.4 -Kv 300 -Cs 2.7 -ExtPhase 90 180

There are examples showing that adding phase shift estimation improves the accuracy of CTF estimation. This is likely due to the inaccurate amplitude contrast used in CTF estimation. The default value is 0.07.

AreTomo2 produces two CTF related files; one is the MRC file storing the power spectrum images embedded with estimated CTFs and the other is a text file storing the fitting results in a compatible format as that of CTFFind4. If “-OutImod” is enabled, a third text file is produced in the “_Imod” subdirectory and conforms to IMod.

To have reasonable CTF estimation at high tilts, AreTomo2 assumes the azimuth angle of astigmatism is constant throughout the entire tilt range. Therefore, the azimuth angle is estimated only on the zero-tilt, which is used as a known constant for other tilts.

Reference

Zheng S, Wolff G, Greenan G, Chen Z, Faas FGA, Bárcena M, Koster AJ, Cheng Y, Agard DA. [AreTomo2: An integrated software package for automated marker-free, motion-corrected cryo-electron tomographic alignment and reconstruction](#). J Struct Biol X. 2022 May 10;6:100068. doi: 10.1016/j.yjsbx.2022.100068. PMID: 35601683; PMCID: PMC9117686.

Technical Support

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